Trophic niche similarity among sea trout *Salmo trutta* in central Norway investigated using different time-integrated trophic tracers

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**ABSTRACT:** From 2011 to 2013, anadromous brown trout *Salmo trutta* (213–730 mm, total body length, *L*₇) were collected during or shortly after their marine feeding migration at 7 different localities in central Norway. The mean volume of stomach content (%) of marine fish prey eaten by *S. trutta* captured in marine waters varied from 34 to 89%. There was a high prevalence (67–100%) for parasite groups potentially transmitted by marine prey fish (i.e. nematodes, cestodes and trematodes) at all sampling sites. There was a significant overlap in the signatures of both δ¹³C and δ¹⁵N in the muscle tissue between the 7 groups of *S. trutta*; however, individual variation within groups was large. A strong positive relationship between δ¹³C and *L*₇ indicated size-dependent niche selection, with smaller individuals feeding less on marine prey and more on brackish or freshwater invertebrates in the estuary. Short-term gut contents data and trophically transmitted parasites showed that all size groups were feeding on marine fish. However, an increased dependence upon marine prey fish by larger *S. trutta* was indicated by a strong positive relationship between *L*₇ and δ¹⁵N. Similarities in *S. trutta* feeding and time-integrated trophic tracers (stable isotopes and parasites) across the 7 localities supports the general view that *S. trutta* feed within similar marine trophic niches. This similarity in feeding niche requirements may make *S. trutta* populations vulnerable to anthropogenic ecosystem perturbations which reduce the diversity of potential marine prey items.

**KEY WORDS:** Feeding ecology · Marine migration · Niche overlap · Brown trout · Stable isotope analyses · Trophically transmitted parasites · Stomach contents

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During the last several decades, the abundance of *S. trutta* has declined markedly in many regions (ICES 2013). As an example, with the exception of northernmost areas, catches in Norwegian rivers have declined by 24–77% during the last 20 yr (Anon. 2015). Recent findings from several other countries (e.g. England, Ireland and Sweden; ICES 2013) indicate similar declines. It has been hypothesised that declines are related to decreased sea survival caused by changes in food supply or increasing infection by the ectoparasitic salmon lice *Lepeophtheirus salmonis* Krøyer, 1838 (ICES 2013, Thorstad et al. 2015).

Anadromous *S. trutta* growth depends largely on the availability of marine prey (Nall 1930, Wootton 1998). Growth rates increase abruptly when anadromous salmonids move from fresh to salt water (Jonsen 1985), probably because of improved feeding opportunities which more than compensate for increases in the energy costs of ionic regulation (McKay & Gjerde 1985). However, knowledge of the marine feeding ecology of *S. trutta* is limited (Rikardsen et al. 2006). This is because *S. trutta* are difficult to catch at sea and most feeding studies are based on small sample sizes (Knutsen et al. 2001). The few studies of *S. trutta* feeding that exist highlight the dietary use of marine prey, such as small fish and large crustaceans (Gronvik & Klemetsen 1987, Elliot 1997, Knutsen et al. 2001, Rikardsen et al. 2007, Knudsen et al. 2011), and seasonal variability of the diet (Rikardsen et al. 2006). In addition, parasite studies have shown *S. trutta* to be infected by generalist parasite species transmitted via amphipods or small prey fish such as gadoids and herring (Hemmingsen & Mackenzie 2001, Knudsen et al. 2011). Nonetheless, there is a paucity of information on general life history characteristics of *S. trutta* (Thorstad et al. 2016), including comparative regional analyses of differences in feeding habits.

In light of limited information on the marine diet of *S. trutta*, the aim of the present study was to describe the marine diets and associated trophic niches of *S. trutta* populations from central Norway. Trophic niches were analysed using stomach content analysis (Hyslop 1980) and 2 time-integrated trophic tracers: trophically transmitted parasites (Knudsen et al. 1996) and stable isotopes (Michener & Kaufman 2007). The use of both tracers provided indirect measurements of long-term trophic behaviour and resource use and made it possible to increase sample sizes by examining marine feeding ecology based on *S. trutta* captured in the sea and in rivers a short time after re-entry from marine feeding. As marine fish seem to be common prey for anadromous *S. trutta*, we hypothesised that *S. trutta* from different populations within a coastal area would have similar marine trophic niches. Further, we hypothesised *S. trutta* would display generalist diets at the population level, but specialized feeding at the individual level, due to size-related feeding preferences.

**MATERIALS AND METHODS**

**Study areas**

*Salmo trutta* was collected from 7 different localities in central Norway, ranging from the Rauma River (62.55653° N, 7.68825° E) in the south to the Drevja River (65.94587° N, 13.13526° E) in the north (Fig. 1). Localities differed in river catchment area (28–3119 km²), the length of the lower river available to anadromous fish (6–42 km), and their distances from the open sea (0–73 km; Table 1).

**Sampling of *S. trutta* and potential prey**

All sampling took place during June–September 2011–2013 (Table 1). In total, 258 *S. trutta* were obtained using different collection methods from 7 localities (5 rivers/river mouths and 2 fjords; Fig. 1, Table 1). Based on subsequent δ³⁴S analyses (see ‘Stable isotope analyses’), 40 individuals were defined as resident trout (non-migratory) and were omitted from further analysis.

In the Drevja, Hundåla and Rauma Rivers, rotenone had been used previously to eradicate *Gyrodactylus salaris* from the watercourses. The aim of the treatment was to kill all carriers of the parasite in the river system. As a part of the programme, dead fish were collected and preserved frozen for future studies. *Salmo trutta* individuals from these water courses were randomly sampled from the central storage facility maintained by the Norwegian Veterinary Institute and used in the present study. The location at Agdenes was not connected to a specific river system and consisted of catches from several local river systems obtained from fish confined to the fjord area with 2 bag nets (mesh sizes 40 and 58 mm) at a research facility located at the mouth of the Trondheim Fjord. In the Nidelva River and the Hopaelva River estuary, *S. trutta* were captured by local game fishers with rod and line, while individuals from the marine Hemnfjord site were captured by gillnets (mesh size 35 mm). Total length (*L*, mm) was measured from the tip of the snout to the tip of the longest
lobe of the caudal fin without compressing the lobes along the midline, and mass (g) was obtained for all individuals. Differences in length and mass of fish between the sampling sites may partly be an artefact of sampling bias introduced by the different collection methods.

Potential prey species (e.g. forage fish and benthic invertebrates) were collected from June to September 2012 by trawling, gill netting and individual sampling in the marine Hemnfjord and then frozen. The samples (n = 40) included Gasterosteus aculeatus, Sprattus sprattus, Spinachia spinachia, Myoxocephalus scorpius, Gobiusculus flavescens, Clupea harengus, Scomber scombrus, Pleuronectiformes sp., Macrobrachium lanchesteri, Crangon crangon, Neomysis integer, Gammarus sp., Euphausiacea, Polychaeta and Mysida.

Analyses of stomach contents

A volumetric analysis of stomach contents was conducted following Hyslop (1980). Stomach contents from the upper end of the oesophagus to the pyloric sphincter were identified to the lowest practical taxonomic level (typically order or family) under a stereoscopic microscope. For most analyses, all orders and families were grouped into ‘surface insects’ or ‘benthic invertebrates’. The relative importance of each prey category was evaluated as volume % for each stomach and the total volume of food category taken by all sampled fish with stomach contents was given as a percentage of the total volume of each stomach content (Hyslop 1980). Gut contents continue to be degraded by stomach acid even if the fish is dead. Consequently, the difference between sampling sites in the ratio of empty stomachs may partly be a result of differences between time of death and gut content sampling at the different locations.

Stable isotope analyses

For stable isotope analyses (δ¹³C, δ¹⁵N, δ³⁴S), a sample of dorsal muscle tissue (e.g. Pinnegar & Polunin 1999) was dissected from all S. trutta and frozen until analysed. An exception was made with muscle tissue samples from the Hopaelva estuary which were preserved in 96% alcohol. Muscle tissue is commonly used to determine long-term diet, and in temperate and northern fish typically

Table 1. Overview of Salmo trutta sampling locations, 2011–2013. NA = no available information

<table>
<thead>
<tr>
<th>Location</th>
<th>Catchment area (km²)</th>
<th>Length of anadromous stretch (km)</th>
<th>Distance to open sea (km)</th>
<th>Sampling place (fresh water/marine water)</th>
<th>Sampling time</th>
<th>Sampling method</th>
</tr>
</thead>
<tbody>
<tr>
<td>Drevja River</td>
<td>178</td>
<td>25</td>
<td>67</td>
<td>River</td>
<td>Aug 2011</td>
<td>Rotenone</td>
</tr>
<tr>
<td>Hundåla River</td>
<td>39</td>
<td>6</td>
<td>58</td>
<td>River</td>
<td>Aug 2011</td>
<td>Rotenone</td>
</tr>
<tr>
<td>Nidelva River</td>
<td>3119</td>
<td>8</td>
<td>73</td>
<td>River</td>
<td>Jun–Sep 2011/2012</td>
<td>Rod and line</td>
</tr>
<tr>
<td>Agdenes</td>
<td>NA</td>
<td>NA</td>
<td>0</td>
<td>Marine</td>
<td>Jun–Aug 2012</td>
<td>Bag net</td>
</tr>
<tr>
<td>Hemnfjord</td>
<td>NA</td>
<td>NA</td>
<td>30</td>
<td>Marine</td>
<td>Jun–Sep 2012</td>
<td>Gillnet</td>
</tr>
<tr>
<td>Hopaelva estuary</td>
<td>28</td>
<td>10</td>
<td>4</td>
<td>Estuary</td>
<td>Jul–Sep 2012</td>
<td>Fly fishing</td>
</tr>
<tr>
<td>Rauma River</td>
<td>1240</td>
<td>42</td>
<td>72</td>
<td>River</td>
<td>Aug 2013</td>
<td>Rotenone</td>
</tr>
</tbody>
</table>

Fig. 1. Location of Salmo trutta sampling sites (2011–2013)
reflects the summer period of somatic growth, with the tissue turnover of muscle depending on growth rate (Trueman et al. 2005). For rapidly growing salmonid fishes, the isotopic value of muscle will equilibrate to diet within the order of a few months (Perga & Gerdeaux 2005, Trueman et al. 2005, Phillips & Eldridge 2006). As a consequence, muscle tissue is considered a useful surrogate for the study of temporally integrated feeding in anadromous salmonid fishes (Doucett et al. 1999b, Etheridge et al. 2008, van der Velden et al. 2012), with δ13C and δ15N stable isotope ratios of a consumer reflecting the isotopic signatures of the prey consumed during the time period that the tissue was synthesised (Fry 2006). The δ13C signature provides an indication of the origin of the carbon source since littoral, pelagic and profundal carbon sources in freshwater have different δ13C signatures, decreasing along littoral–profundal gradients (Van der Zanden & Rasmussen 1999). In the marine environment, the δ13C value of marine carbonates (dissolved inorganic carbon) tends toward 0‰ with the result that the δ13C of resident biota are also elevated (Sharp 2007). In contrast, the δ15N signature of an organism indicates trophic position, since δ15N signatures increase with an increase in trophic level (Van der Zanden & Rasmussen 1999, Post 2002). Further, the stable signature ratio of δ34S reflects whether individuals have previously been to sea (Doucett et al. 1999a), and coupled with δ13C, may be used to screen organisms for marine residency.

Here, we sorted captured fish into anadromous and non-anadromous groupings based initially on their δ13C value, with all fish having δ13C values > −22‰, being treated as anadromous given that δ13C = −22‰ is the lower δ13C limit for marine organisms found in central Norwegian coastal waters (Fredriksen 2003). The criterion was considered conservative and likely to result in the inclusion of both anadromous and non-anadromous fish in a single group. Accordingly, fish with δ13C values < −22‰ were re-analysed using δ34S to more precisely establish anadromy, with higher δ34S values (>10‰) being considered indicative of anadromy (Doucett et al. 1999a).

Tissues from both S. trutta and potential prey were dried at 50°C for 24 h. At the University of Waterloo, Canada, the dried tissue was ground to a fine powder with a mortar and pestle and analysed for stable isotope ratios (δ13C, δ15N, δ34S) using the methods described by Guiguer et al. (2002), Power et al. (2009) and van der Velden et al. (2012). Analyses for δ13C and δ15N were performed using a Delta Plus Continuous Flow Stable Isotope Ratio Mass Spectrometer (Thermo Finnigan) coupled to a Carlo Erba Elemental Analyzer (CHNS-O EA1108, Carlo Erba). Analysis of δ34S was completed using an Isochrom Continuous Flow Stable Isotope Ratio Mass Spectrometer (GV Instruments, Micromass) coupled to a Costech Elemental Analyzer (CNSO 4010, Costech Analytical Technologies). Working internal laboratory standards were calibrated against the International Atomic Energy Agency standards CH6 for carbon, N1 and N2 for nitrogen, and SO-5, S1 and S2 for sulphur and were run as controls throughout the analysis to ensure the continued accuracy of all measurements (±0.2‰ for carbon, ±0.3‰ for nitrogen and ±0.5‰ for sulphur in an organic material). Analytical precision was assessed by mean differences of 1 in 10 duplicate samples, where the mean ± SD was 0.14 ± 0.2‰ for δ13C and 0.18 ± 0.2‰ for δ15N. All results were expressed in standard δ notation (Perga & Gerdeaux 2005).

The C:N ratios of studied fish were uniformly low (C:N < 4 in 94% of samples), which suggests carbon isotope signatures can be left uncorrected for lipids (e.g. Post et al. 2007, Jardine et al. 2013). Use of this rule is unlikely to bias analytical results as individuals with C:N > 4 were drawn from 6 of the studied populations and from sizes in the range of 272–730 mm. Furthermore, as noted by Fagan et al. (2011), arbitrary use of lipid correction is itself questionable, given the demonstrated lack of relationship between lipid-corrected values and actual measured lipid levels in fish. Given the limited need for lipid correction based on the <4 rule, the random occurrence of individuals with a high C:N ratio within and among populations and the published evidence questioning the use of routine lipid correction in fish ecological studies, the bulk carbon isotope values used in this study were not lipid-corrected.

The need for baseline adjustments was assessed using 1-way ANOVA followed by Tukey’s post-hoc honestly significant difference test (e.g. Zar 2009). Such an approach works well if the δ15N values of the assumed primary consumers are near 0‰ (e.g. Hobson et al. 2002, Jennings & Warr 2003, Søreide et al. 2006), as they were here. Stable isotope data (δ15N) for available lower trophic level (e.g. Amphipoda, Mysida, Mytilus edulis) invertebrates sampled from across the latitudinal range 62.3–63.3°N along the central Norwegian coastal area proximate to the S. trutta sampling sites were not significantly different (ANOVA, $F_{2,30} = 2.566, p = 0.094$). Although the tested range encompassed only a portion of the range for which S. trutta samples were available (63.0–65.3°N), the data available for testing did not support the use
Analyses of trophically transmitted parasites

Marine parasites of S. trutta were recorded and enumerated from the available frozen intestine and stomach of individual fish as a standard method. Based on a salmonid study from a northern Norwegian fjord system (see Knudsen et al. 2011), parasites were divided into 2 groups according to their intermediate host type (MacKenzie & Abaunza 1998): (1) parasites transmitted by gammarids, such as the acanthocephalan Echinorhyncus gadi (Zoega, 1776), and (2) parasites transmitted mainly by marine fish, such as trematodes (e.g. Lecithaster gibbosus Rudolfi, 1802 and Derogenes varicus Müller, 1784), and several nematode species and adult cestodes (Eubothrium sp. Nybelin, 1922). The terms ‘prevalence’ (i.e. the proportion of infected hosts) and ‘abundance’ (i.e. the mean number of parasites in both infected and uninfected hosts) sensu Bush et al. (1997) were used here to describe parasite occurrences in S. trutta.

Data analyses

Prey δ¹⁵N and δ¹³C signatures were corrected for trophic enrichment (Δ) using, respectively, mean fractionation factors of 3.23 and 1.03, and are presented as post-fractionation equivalents (plotted prey δ values = prey isotope values + Δ) when compared or plotted with S. trutta stable isotope values (e.g. Jensen et al. 2012).

The SIBER package (Stable Isotope Bayesian Ellipses in R, version 2.0.3, Jackson & Parnell 2016) was used to study isotopic niche widths. Preservation of muscle tissue in 96% alcohol may influence δ¹⁵N and δ¹³C signatures and confound correlations with signatures from frozen material (Stallings et al. 2015). Consequently, the samples from the Hopaelva estuary were omitted from the analyses of isotopic niche widths.

To analyse if either δ¹³C or δ¹⁵N were dependent on the fish L̄, a general linear model (GLM) with a Gaussian error distribution and identity link function were used. Collection site was included as a grouping variable to compare the strength of ontogenetic shifts among populations. The possible relationship between L̄ and the tendency to be piscivorous was tested by a binomial logistic regression model using the logit link. All statistical analyses were conducted using RStudio version 1.0.44 (www.rstudio.com).

RESULTS

From 258 Salmo trutta from the 7 locations (Fig. 1), 218 individuals (213–730 mm L̄, Table 2) were identified as being anadromous using stable isotope data. The samples from the Rauma River had the largest mean L̄ and mass (599 mm, 2292 g), while S. trutta with the smallest mean L̄ and mass were from the Drevja River (313 mm, 364 g).

Stomach contents

S. trutta captured in marine waters (Agdenes, Hemnfjord and Hopaelva estuary) in general had higher amounts of marine fish in their stomachs than individuals sampled in the rivers. The mean volume (%) of prey fish in individuals captured in the Hopaelva estuary and in the Agdenes bag nets in the

<table>
<thead>
<tr>
<th>Location</th>
<th>n</th>
<th>Total body length (mm)</th>
<th>Empty stomachs (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Mean (SD)</td>
<td>Range</td>
</tr>
<tr>
<td>Drevja River</td>
<td>28</td>
<td>316 (62)</td>
<td>218−442</td>
</tr>
<tr>
<td>Hundåla River</td>
<td>80</td>
<td>367 (80)</td>
<td>213−583</td>
</tr>
<tr>
<td>Nidelva River</td>
<td>6 (4*)</td>
<td>522 (167)</td>
<td>347−730</td>
</tr>
<tr>
<td>Agdenes</td>
<td>37</td>
<td>433 (77)</td>
<td>346−660</td>
</tr>
<tr>
<td>Hemnfjord</td>
<td>32</td>
<td>356 (72)</td>
<td>235−495</td>
</tr>
<tr>
<td>Hopaelva estuary</td>
<td>21</td>
<td>369 (85)</td>
<td>262−600</td>
</tr>
<tr>
<td>Rauma River</td>
<td>14</td>
<td>599 (82)</td>
<td>485−730</td>
</tr>
</tbody>
</table>

*Number of fish used in calculation of mass
fjord was 89% (Fig. 2). Marine-captured *S. trutta* from Hemnfjord averaged 34%, while individuals sampled in the rivers after the marine feeding migration never had more than 18% fish content in the stomach. Instead, most of the identified stomach contents consisted of benthic invertebrates and surface insects (Fig. 2). Identified marine fish prey consisted of the pelagic fish species sprat *Sprattus sprattus* L., herring *Clupea harengus* L. and small sandeel *Ammodytes tobianus* L. The proportion of empty stomachs (Table 2) ranged from 21% (Rauma River, n = 14) to 100% (Nidelva River, n = 6).

The prey fish described above were found in the stomach contents of all 3 size groups (Fig. 3), and there was no increase in the percent fish content with size (*r*² = 0.004, *n* = 70, *p* > 0.05). Surface insects were found in all length classes, and there was no correlation between percent volume and size of the *S. trutta* (*r*² = 0.0006, *n* = 70, *p* > 0.05). *Gammarus* spp. were found only in fish >300 mm.

Stable isotope signatures

There was a large overlap in the SIBER ellipses between *S. trutta* from the 6 localities (Fig. 4), with a number of the ellipses for populations being wholly contained in the ellipses of other populations. In total, 90% of the *S. trutta* had values for δ¹³C and δ¹⁵N within the ranges of −18.7 to −25.2‰ and 7.9 to 14.5‰, respectively. The δ¹³C and δ¹⁵N signatures from *S. trutta* and their potential prey in the marine Hemnfjord indicated that they had mainly been feeding on krill and pelagic marine fishes (Fig. 5).

Overall, there was an increase in the δ¹³C signature (GLM, *n* = 197, *F* = 36.179, *p* < 0.001) and δ¹⁵N signature (*n* = 197, *F* = 94.102, *p* < 0.001) with increasing *L*_T. The strength of the correlation differed between collection sites (δ¹³C signature: *n* = 7, *F* = 9.843, *p* < 0.001; δ¹⁵N signature: *n* = 7, *F* = 33.609, *p* = 0.002). *S. trutta* from Agdenes and the Rauma River did not show any relationship between δ¹³C and *L*_T (Agdenes: *n* = 37, *F* = 2.763, *p* = 0.11; Rauma River: *n* = 14, *F* = 0.491, *p* > 0.05), while *S. trutta* from the Drevja and Nidelva Rivers did not show any relationship between δ¹³C or δ¹⁵N signatures and *L*_T (δ¹³C:...
Davidsen et al.: Trophic niche similarity among sea trout

Composition of trophically transmitted parasites

There was a high prevalence (67−100%) of the parasite groups potentially transmitted by marine fish as prey at all sampling sites, for *S. trutta* caught both in the river and in the sea (Table 3). Cestodes, mainly marine *Eubothrium* sp., were the most prevalent (45−92%). Trematodes varied more (8−80%) and nematodes ranged from no infection (river samples from Rauma) up to 54% (marine samples from Agdenes). The abundance of parasite groups potentially transmitted by fish was low to intermediate and varied from 1.8 (river samples from the Hundåla River) up to 13.3 (marine samples from the Hopaelva estuary). However, there were no differences in levels of prevalence (t-test; n = 8, p > 0.05) or abundance (n = 8, p > 0.05) between individuals captured in marine water or freshwater. The infection level of Acanthocephala, which have marine crustaceans as an intermediate host, was generally low as indicated by prevalence (highest 15% in Drevja River samples) and abundance (highest 1.4 in marine samples from Hemnfjord). The smallest piscivorous *S. trutta* was 213 mm based on the abundance of parasite groups potentially transmitted by fish. There was no relationship between $L_T$ and the tendency to be a piscivore (logistic regression, n = 156, p > 0.05) based on stomach content or trophically transmitted parasites.

**DISCUSSION**

This study documented the regional similarity of *Salmo trutta* population marine trophic niches as
measured by stable isotope and marine parasite tracers along a 400 km coastal area of central Norway. Stomach contents from individuals that were caught in marine water consisted mainly of marine fish, while the stomachs of individuals captured in the rivers had higher abundances of benthic invertebrates and surface insects. The abundance and prevalence of parasite groups potentially transmitted by marine fish did not differ between individuals captured in marine water or in the rivers, indicating that _S. trutta_ captured in the river had also been mainly feeding on marine fish sometime prior to their capture. Likewise, stable isotope analyses confirmed similar resource utilisation across the sea-migratory populations. Our study indicated a diverse, habitat-specific diet of _S. trutta_ which concurs with earlier characterisations of _S. trutta_ as an opportunistic feeder (Pemberton 1976, Knutsen et al. 2001, Rikardsen et al. 2006), although there is an apparent preference and reliance on smaller pelagic fishes during their marine feeding migration.

Analyses of stomach contents showed that _S. trutta_ feed on a variety of prey items, but fish were the dominant prey found in individuals captured in marine waters (Agdenes, Hemnfjord and Hopaelva estuaries). Reliance on fish as prey has also been documented in earlier studies of _S. trutta_ diets (Grønvik & Klemetsen 1987, Knutsen et al. 2001, Knudsen et al. 2011), and for other coastal feeding anadromous salmonids such as Arctic char _Salvelinus alpinus_ (L.) (Klemetsen et al. 2003, Rikardsen & Dempson 2011) and Atlantic salmon _Salmo salar_ (Renkawitz et al. 2015). The occurrence of surface insects and benthic invertebrates in stomachs from _S. trutta_ sampled in the rivers after the marine feeding migration indicates that these _S. trutta_ also feed in the river mouth or in the river after returning from the sea.

Stable isotope analysis of muscle tissue from the _S. trutta_ in this study revealed a broad range of signatures for both δ¹³C and δ¹⁵N, although the values tended toward the higher values indicative of marine feeding (e.g. Doucett et al. 1999a, Etheridge et al. 2008). The range of δ¹³C indicates that the _S. trutta_ have a mixed feeding strategy, with comparison of stable isotope signatures from _S. trutta_ and their potential prey in Hemnfjord indicating mixed feeding on krill, and pelagic and littoral fishes. Individuals with δ¹³C values in the range of −21 to −22‰ may only have fed for a short time in the marine habitat or may have mainly preyed upon brackish or freshwater invertebrates in the estuary (Etheridge et al. 2008). While the majority of stable isotope signatures are consistent with marine feeding, differences were evident in the stomach content analyses among sites. These results suggest rapid transition from fish to invertebrate prey as _S. trutta_ re-enter freshwater. Lags in tissue turnover in salmonids are typically on the order of months (Trueman et al. 2005), implying that despite evidence of recent freshwater feeding and residency, the majority of piscivorous _S. trutta_ sampled in this study reared in the marine environment.

The observed relationship between _L_T_ and δ¹⁵N is indicative of size-dependent feeding strategies, with smaller _S. trutta_ feeding on a mixture of invertebrate and fish prey in estuaries and larger _S. trutta_ feeding mainly on marine prey. Accordingly, there are ontogenetic shifts in prey selection (e.g. Guiguer et al. 2002, van der Velden et al. 2012), with larger individuals feeding at higher trophic levels because of increased forage fish consumption. That no such relationship was found in the _S. trutta_ from the Drevja and Nidelva Rivers is likely due to the low mean _L_T_ (Drevja) and low number of sampled fish (Nidelva).

Marine parasites were frequent (67–100%) on _S. trutta_ at all sampled localities, and infection patterns (prevalence and abundance) showed consistency across different habitat types (i.e. fjords and river systems) across the entire geographic region. Parasite species potentially transmitted by fish as prey (i.e. nematodes, cestodes and trematodes) had a much higher prevalence than parasites transmitted by crustaceans (i.e. Acanthocephala), which may be because fish are the main dietary component of _S. trutta_. Also, _S. trutta_ caught in the sea had similar patterns of infection to those individuals caught in the river, suggesting that prevalence of marine parasites is a good indicator of anadromous life-history strategies in general, both at the individual (Knudsen et al. 2011, 2014) and the population level (e.g. MacKenzie & Abaunza 1998, Knudsen et al. 2005).

Generally, the endoparasites recorded from central Norwegian _S. trutta_ had lower abundance compared to those recorded from northern Norway (Knudsen et al. 2011). Parasite infections have been shown to impair swimming performance and lead to decreases in burst swimming speed, migration performance and fatigue distance (Barber & Poulin 2002). High parasite burdens have also been associated with pale gills, decreased body condition and host mortality in farmed _S. trutta_ (Rubio-Godoy & Tinsley 2008), with high loads of intestinal helminths in particular seen as the probable trigger for long-term malnutrition leading to increased _S. trutta_ emaciation and mortality (Mladineo et al. 2009). High parasite loads may also affect fish behaviour, altering habitat use and foraging opportunity as infected individuals adopt
movement patterns expected to suppress infestation rates (Gjelland et al. 2014). Lowered levels of parasites observed at sites sampled here suggest reduced parasite-induced energetic and fitness costs for S. trutta in central Norway. A possible exception may be those individuals that had relatively high densities of the adult cestode, Eubothrium sp. This genus has been shown to have negative consequences for individuals’ somatic growth (Bristow & Berland 1991, Saksvik et al. 2001).

Examination of stomach contents indicated that small pelagic fish species like sprat Sprattus sprattus, herring Clupea harengus and small sandeel Ammodytes tobianus are important parts of S. trutta diets, as has been similarly noted in the studies of Knutsen et al. (2001) and Rikardsen & Amundsen (2005). Observed trophic niche similarity across sea trout populations over a large geographical area suggests that sea trout may be vulnerable to fluctuations in these prey populations.

In partially anadromous species, marine migrations are not obligate and individual fish may change life history tactics (i.e. anadromy versus freshwater residency) to maximize fitness if environmental conditions change (e.g. Nordeng 1983, Jonsson & Jonsson 1993). Indeed, anadromy may have evolved in the first place as a response to situations where food resources in the sea exceeded those in freshwater (Gross 1987). Thus, significant decreases in biomass and reduced availability of pelagic prey fish species for larger S. trutta may trigger alterations in their feeding behaviour by reduced feeding time at sea and/or shifting diet to a heavier reliance on littoral marine prey, or even a change in life history tactics to feed exclusive on freshwater prey resources.

Other preferred dietary items in addition to the species mentioned above may include fishes of the families Clupeidae and Gobiidae in areas and years when they are abundant (Knutsen et al. 2001, 2004), but these were not observed here. Prey fish species found in this study have generally been subject to wide fluctuations in relative abundance. For example, the biomass of A. tobianus has declined markedly in the last 2 decades (ICES 2015). Likewise, C. harengus experienced a significant decline in abundance in the 1970s (ICES 2015). Fluctuations in the relative abundances of small prey fishes have been correlated with size (mass) of other coastal feeding salmonids such as Salvelinus alpinus (Dempson et al. 2002).

The similarities in marine-captured S. trutta diets and stable isotope signatures and the pattern of infection of marine parasites suggests that there is rough equivalence in the foraging niches across S. trutta populations along the coastal area of central Norway. Results thus indicate that the marine environment in the actual geographical region has relatively similar diet resources for anadromous salmonid populations. The geographic equivalence of the trophic niche of anadromous S. trutta could imply that they are vulnerable to possible future changes in the coastal environments that would affect the densities of favoured marine prey. Thus, the variable density of the marine prey fish species used by S. trutta along the coast could explain some of the declines in the recorded densities of sea trout populations that have been observed in recent decades (ICES 2013, Anon. 2015). As S. trutta in general appear to depend on pelagic fish species such as S. sprattus, C. harengus and A. tobianus, special attention needs to be given to the conservation of these species as they represent important prey resources for S. trutta.

In conclusion, the large similarity observed in S. trutta feeding niches from the 7 localities found using stomach contents combined with 2 time-integrated trophic tracers (trophically transmitted parasites and stable isotopes) suggests that S. trutta occupy similar trophic niches at broad regional scales. While it was found that the S. trutta have generalist diets at the population level, as individuals they tend to specialize due to size-related feeding preferences. Such specialization may make S. trutta vulnerable to anthropogenic ecosystem perturbations (e.g. climate change, overfishing, shoreline development) that may change the diversity of potential marine prey items and thus cause a reduction in their abundance.

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LITERATURE CITED


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